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Functions and Interrelationships of Leukocytes in Inflammation as Elucidated by the Rebuck Skin Window Technique

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This article provides an overview of the functions of human leukocytes in inflammation as elucidated by the Rebuck skin window technique. The migration sequence of various leukocytes into the field of inflammation is described, as well as cytologic, cytochemical, and transformational changes, and the interrelationships of responding leuko-

cytes. Since it was introduced in 1955, the Rebuck skin window technique has provided an excellent means of studying in vivo inflammatory response to different phlogistic agents in normal individuals and in various disease states. This simple technique continues to prove fruitful to further study and to monitoring of many disease states.

Since detailed descriptions of the stages of inflammation can be found in numerous texts (1-6), this article will present an overview of leukocytic migration, morphologic changes, and functions as elucidated by the Rebuck skin window technique (7-9) in normal subjects as well as in various disease states.

The classic article by Rebuck and Crowley (7) describing their simple yet definitive method of serially studying the exudative response to phlogistic agents is reproduced in this section (pp. 184-209). Since its publication, this reproducible and inexpensive method has been extensively used (9). It has allowed monitoring of leukocytic reactions and functions in response to many different inflammatory excitants in the normal individual as well as in several disease states. Over the years, it has undergone few alterations and, if anything, has become simpler to use because of the availability of chemically and antigenically inert sterile plastic cover slips, hypoallergenic adhesive tape, etc. Also, modifications of the scarification technique and semiautomated mechanical means of scarification have been introduced (10-16) that may provide standardization in the hands of the inexperienced. However, the original scarification procedure is quite easy to learn and reproduce with minimal experience.

Mechanical trauma produced by scarification alone could be expected to produce hyperemia and prestasis as factors for increasing vascular permeability (7,18). In most types of injury, biphasic permeability has been demonstrated (19-21); an early but transient phase is followed by a variable latent period and a late prolonged phase of increased permeability. The increase in permeability is due mainly to histamine release (22) and to the action of other vasoactive peptides and kinins (23-38). Depending upon the type, extent, and severity of the injury, the amount of plasma and leukocyte migration through the gaps between the endothelial cells (39,40) varies considerably. However, the earliest responding cells are almost always polymorphonuclear neutrophilic granulocytes (PMNs) (21,-30,31). Their emigration by active ameboid movement from the intravascular areas to the site of inflammation has been fairly well established (1,40-51). The skin window technique has repeatedly substantiated PMN migration as an early event (7,8,52). At the same time it is important to remember that the trauma of the technique alone produces only sparse leukocytic response consisting of a few neutrophils, a rare lymphocyte, a few local tissue macrophages, and a few hematogenous monocytes.

Leukocytic Response to Antigenic Exposure

The inflammatory response to an antigen in an individual without prior exposure should be related to its local phlogistic characteristics without the mediation of antigenantibody reactions, complement activation, etc. Hurley

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Reaction to Trauma of the Technique

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(40) has shown that injecting inert substances produces no increase in vascular permeability but causes delayed leukocytic migration. However, injecting histamine or serum causes both increased vascular permeability and delayed cell migration. The skin window technique demonstrates that applying diptheria-tetanus toxoid (DT 0.5m, Lilley) to the scarified skin of nonimmunized individuals induces cell migration and changes comparable to what could be expected by the mechanical trauma of the technique alone (9). However, since most individuals in the U.S. are immunized to DT, dynamic leukocytic response is elicited when DT is applied to the scarified area of most individuals.

Many articles by Rebuck and others (7,8,52) have provided descriptions of the leukocyte types, numbers, morphologic changes, etc. Recently, Sokol, Durrant, and Hudson (53) have presented information gained by scanning electron microscopy of skin window cells from normal subjects. In brief, the first wave of emigrating cells are PMNs, blood monocytes, and occasional lymphocytes. The number of emigrating monocytes is generally comparable to the numbers encountered in the blood stream. Although the number of emigrating PMNs decreases with the timing of the sample in 6-24 hour cover slip preparations, they show dynamic cytoplasmic and nuclear changes. PMN nuclei become progressively hypersegmented with 1.3% of the cells having more than five lobes or up to ten at 10-12 hours of response. In the cytoplasm, glycogen synthesis increases and is later transferred to mononuclears for energization. Lipid content of the migrated PMNs is high, and most enzymatic reactions of PMNs continue to be enhanced until they lose their viability (7-9). The only known enzyme to diminish quickly in the PMNs is leukocyte alkaline phosphatase.

As mentioned, monocytes responding to inflammation are numerically equivalent to the numbers (percentages) in the blood (9). All authorities agree that blood monocytes are capable of transforming into macrophages at the inflammatory site (9,52,55). The 3-12 hour cover slip preparations have shown that the monocytes are metallophilic, concentrate dyes, and progressively demonstrate the cytochemical features of macrophages (8). Recent experiments indicate that mononuclear cells leave the vessels more or less simultaneously with PMN leukocytes but persist longer in the exudate because of their longer life span (56-58). Furthermore, macrophages originate predominantly from blood monocytes and not from tissue histiocytes (57,59).

The role of lymphocytes in acute inflammatory response and their capability to transform into macrophages is still fervently debated (7,8,52,56,57,60). Originally, Metchnikoff observed (61) that most macrophages are transformed lymphocytes. Experiments by Kolouch (62), Townsend and Campbell (63), Good (64), Berman (65), and many others

(79,135) have provided conclusive support for lymphocyte to macrophage transformation as reported by Rebuck and Crowley (7). Subsequent application of the skin window technique in experimental inflammatory lesions in man has yielded confirmatory evidence that human lymphocytes are indeed capable of transforming into macrophages (55. 67-74). At three hours the emigrated lymphocytes in the skin windows are scanty. However, 9-14 hour preparations reveal that most of the responding mononuclear cells are small and medium-sized lymphocytes that slowly hypertrophy by increase in colorless cytoplasm, as well as by basophilic material in the enlarging cell body. An increase in the irregularity of the nuclear membrane and loosening of dense chromatin accompany hypertrophy. At 14-18 hours, further lymphocyte hypertrophy and other transformational changes lead to the formation of lymphocytogenous macrophages. Cytochemical analysis of the transformational process has shown that in addition to the presence of acid phosphatase and glycogen, which have been noted in the circulating lymphocyte, the transforming lymphocytes acquire such enzymes as oxidase, peroxidase, alkaline phosphatase, as well as sudanaphilic cytoplasmic constituents (9). These lymphocytes also become phagocytic for vital dyes, pyrrol blue, lithium-carmine, trypan blue and trypan red (8).

Improved knowledge of the lymphocyte physiology and functions allows us to recognize lymphocytes as B, T, null, and/or one of the subpopulations. It would appear natural that the T lymphocytes involved in cell-mediated immunity are the ones which would predominantly emigrate to inflammatory sites and then transform to macrophages. It has been shown that applying antithymocyte globulin (ALG) at the 10th hour of an antigenically stimulated skin window site completely eliminates the main mass of small round mononuclears by the succeeding 12th hour of inflammation (9). This not only confirms that most of the responding cells are T lymphocytes but also helps to differentiate further the lymphocytic from the monocytic source of macrophages.

Leukocytic Response in Disease States

Because it allows one to serially sample the exudative response to inflammatory stimuli, the skin window technique has provided an excellent means of studying leukocyte response in many disease states.

Decreased granulocyte emigration and/or function

Decreased emigration of PMNs in skin windows of patients with neutropenia due to varying causes was first reported by Riis (67). The close correlation he found between leukocytic response in skin windows and the inflammatory lesions in the organs of some autopsy patients has been

confirmed by Rebuck, et al (54). It is obvious that delayed or absent PMN response in windows has considerable significance when the capabilities of cellular defenses in disease states are to be correlated. Decreased granulocyte migration is also critical, since initial neutrophilic migration is an essential step leading to the migration of lymphocytes and their successful transformation to macrophages. Impaired neutrophilic response in skin windows has been reported in congenital neutropenia (75), neutrophil actin dysfunction (76), diabetes mellitus (77-79), paroxysmal nocturnal hemoglobinuria (54,80), Chediak-Higashi anomaly (81,82), complement component deficiences (83-85), leukemias and myeloproliferative disorders (9), established malignant states (9), alcoholism (86), burns (87,88), kala azar and Schistosomiasis mansonii (89), and conditions causing elevated serum levels of chemotactic factor inactivator (90). Decreased neutrophilic migration has also been observed in patients who are being treated with ACTH, hydrocortisone, progestron, prednisone, gold salts, chloropheniramine maleate, and imuran (9). Thus, neutropenic individuals are more susceptible to infections and show poor inflammatory response, but other conditions which impair PMN migration, release of hydrolytic enzymes (Metchnikoff's cytases) (61,91,92), and impairment of phagocytosis can also lead to reduced chemotaxis or transformation of mononuclears (48).

Skin window studies have revealed diminished phagocytic capabilities of neutrophils in Hegglins and Pelger Huet anomalies (93), paroxysmal nocturnal hemoglobinuria (54,80,93), chronic granulocytic leukemia (94), uveitis (95), during hydrocortisone or ACTH therapy (96), and when urea, urine (97-99), or crystals of gouty tophi (100) are used in skin windows.

Diseases associated with exuberant granulocyte emigration

An orderly, controlled response by blood and tissue elements is obviously necessary for proper healing of all injuries. Excessive cellular outpouring into the field of inflammation due to a chemotactic or phlogistic agent can be undesirable, as observed in the Schwartzman and the Arthus phenomena (101-103). Interaction of immune aggregates and the complement system releases substances that are strongly chemotactic for PMNs (104-106). Phagocytosis of immune aggregates leads to explosive release of lysosomal material with attendant hydrolysis and other changes (107-112). In allergic individuals, the antigen-antibody complexes are not only chemotactic for PMNs, but are also specifically chemotactic for eosinophils (113-116). It is hypothesized that eosinophils are attracted by allergic responses that promote fibrin formation since the specific eosinophil granules have been shown to contain profibrinolysin.

In the skin windows, excessive or persistent PMN migration has been noted in a few disease states. In patients with nonspecific uveitis, Hessburg and Rebuck (95) found that excessive persistent neutrophilic migration was not followed by normal mononuclear transformations. In these patients, uveal pigment placed at the test sites also resulted in excessive outpouring of eosinophils. In patients with untreated polycythemia vera, increased PMN emigration but decreased mononuclear emigration has also been observed (117).

When Waldmann, et al (118-119) discovered the Fitzgerald factor, they applied the skin window lesion to Mr. Fitzgerald, who lacked this coagulation factor (and after whom it was named). They observed a massive outpouring of PMNs which persisted through the 49th hour of study. Increased numbers of eosinophilic and basophilic granulocytes were also observed at various test intervals. They speculated that the exudative changes in this otherwise healthy individual compensated for the failure of surfacemediated generation of fibrinolytic activity.

While eosinophilic granulocytes normally comprise less than 0.1% of responding cells in the skin windows, markedly increased response in hypersensitive individuals has been reported using many different experiments (9). It has been described in antigen hypersensitivity of immediate type (120) but not in patients with delayed onset of food hypersensitivity. Excessive eosinophil migrations have been induced in windows of challenged, passively sensitized individuals, and Fowler and Lowell (113) have been able to correlate the eosinophilic response to clinical severity. The application of topical steroids fails to abolish eosinophilic response but oral steroid therapy, especially if administered daily, prevents it.

There are not many recognized diseases that affect basophil emigration or their function in inflammation. In a normal control window site the number of emigrating basophils is usually negligible, although the technique has focused specific attention on some disorders of basophilic granulocyte emigration and function. Basophilic granulocytes are the blood-borne counterpart of tissue mast cells and, like them, are a rich source of histamine, heparin, and chymase. Using the skin window technique, Priest and his associates (54,55,121-24) reported basophilic granulocyte outpouring in ulcerative colitis and postulated that the colonic and other lesions were due to abnormal basophil activity and release of excess histamine, heparin, and chymase. This concept is well supported by the fact that there is a significant increase of metachromatically granulated cells in the colonic lesions of ulcerative colitis. Similar abnormal basophilic migration has been seen in the windows of patients with interstitial cystitis (9,123), another disease with a high number of metachromatically-granulated cells in the bladder wall. Abnormal basophil emigration has also been described in patients with furunculosis (125) and in histoincompatibility (126-28).

Diseases affecting monocyte response

Not too many conditions are known to cause lack of hematogenous monocyte response in skin windows. Even in immunosuppressed individuals whose granulocytic and lymphocytic response is below normal, the monocyte response remains reasonably constant unless oversuppression occurs. In chronic immunosuppression, whether induced, acquired or congenital, the monocyte response is increased in proportion to their increase in blood (9).

A very interesting and significant finding in the skin windows of patients with genetic mucopolysaccharidosis is the appearance of increased basophils and quasi-mast cells (123,129-31) due to the ingestion of metachromatic acid mucopolysaccharides by the macrophages. Since metachromatic quasi-mast cell macrophages regularly appear in the skin window, even in forme fruste cases (129), this technique could be helpful in evaluating suspected cases of genetic mucopolysaccharidosis.

Diseases affecting lymphocyte response

Since it is well established that neutrophilic appearance in normal numbers is essential for the emigration, energization, and transformation of lymphocytes to macrophages (9), any disorders which impair neutrophil emigration and release of lymphochemotactic substances will lead to decreased lymphocyte participation and function. For example, in the skin windows of patients with far advanced carcinomas, sarcomas, and Hodgkin's disease decreased lymphocyte response has been described (9).

Perhaps the lymphocytolytic and immunosuppressive agents, such as ACTH, cortisone, imuran, and other antimetabolites not only decrease lymphocyte participation by systemic action but also, as detailed above, by affecting the neutrophil emigration. On the other hand, rapid transformation of lymphocytes to macrophages in Boeck's sarcoid has been reported (9,132), although the exact cause of this rapid transformation is not apparent. It would appear that the disorders specifically involving T lymphocytes should affect the response of lymphocytes and their transformation in skin windows. Depressed cell-mediated immunity and decreased mononuclear response in skin windows have been observed in Darier's disease (133). However, many controlled studies for T cell function, migration, and transformation need to be performed and may provide a fruitful avenue of further research.

References

- Florey HW. Inflammation. In: Florey H, ed. General pathology. 3rd ed. Philadelphia: WB Saunders, 1962.
- Zweifach BW, Grant L, McCluskey RS, eds. The inflammatory process. New York: Academic Press, 1965.
- Hurley JV. Acute inflammation. Baltimore: Williams and Wilkins Co, 1972.
- Thomas L, Uhr JW, Grant L, eds. International symposium of injury, inflammation and immunity. Baltimore: Williams and Wilkins Co, 1964.
- Movat HZ, ed. Inflammation, immunity and hypersensitivity. 3rd ed. New York: Harper and Row, 1978.
- 6. Van Arman CG, ed. White cells in inflammation. Springfield, II: Charles C. Thomas, 1974.
- Rebuck JW, Crowley JH. A method of studying leukocytic functions in vivo. Ann NY Acad Sci 1955;59:757-805.
- Rebuck JW, Boyd CB, Riddle JM. Skin windows and the action of the reticuloendothelial system in man. Ann NY Acad Sci 1960;88:30-42.

- Rebuck JW, Kelley AP, Sweet LC. In: Rebuck JW, Berard CW, Abell MR, eds. Monitoring leukocyte reticuloendothelial system functions in man. IAP Monograph, No. 16. Baltimore: Williams and Wilkins Co, 1975.
- Kiistala U, Mustakallio KK. Dermo-epidermal separation with suction. Electron microscopic and histochemical study of initial events of blistering on human skin window. J Invest Dermatol 1967;48:466.
- 11. Senn HJ. Infekkabwehr bei hamoblastosen: Funktionelle untersuchungen uber leukocytenmobilisation beim gesunden und beim krauken menschen. Vol. 6. Berlin: Springer-Verlag, 1972.
- 12. Ghosh ML, Hudson G, Blackburn EK. Skin window macrophages in malignant lymphomas. Br J Haematol 1973;25:293.
- 13. Mass MF, Dean PB, Weston WL, Humbert JR. Leukocyte migration in vivo: A new method of study. J Clin Lab Med 1975;6:1040.
- Hellum KB, Solberg CO. Human leukocyte migration: Studies with an improved skin chamber technique. Acta Path Microbiol Scand (Sect C) 1977;85:413.

Leukocytes in Inflammation

- Park HB, Dolen J, Snyder B. Defective chemotactic migration of polymorphonuclear leukocytes in Pelger-Huet anomaly. Proc Soc Exp Biol Med 1977;155:51.
- Turnbull LW, Evans DP, Kay AB. Human eosinophils, acetic tetrapeptides (ECF-A) and histamine: Interactions in vitro and in vivo. Immunology 1977;32:57.
- Ricker G. Pathologie als naturwissenschaft: Relationspathologie. Berlin: Springer-Verlag, 1924.
- 18. Krogh A. The anatomy and physiology of blood capillaries. New Haven: Yale University Press, 1929.
- 19. Sevitt S. Local blood-flow changes in experimental burns. J Pathol Bact 1949;61:427.
- Sevitt S. Inflammatory changes in burned skin: Reversible and irreversible effects and their pathogenesis. In: Thomas L, Uhr JW, Grant L, eds. Injury, inflammation and immunity. Baltimore: Williams and Wilkins Co, 1964.
- Burke JF, Miles AA. The sequence of vascular events in early infective inflammation. J Pathol Bact 1958;76:1.
- 22. Feldberg W. Distribution of histamine in the body. In: Wholstenholme GEW, O'Connor CM, eds. Ciba foundation symposium on histamine. London: Churchill, 1956.
- 23. Lewis GP. Natural and synthetic bradykinin. Nature 1960;188:999.
- 24. Lewis GP. Plasma kinins and other vaso-active compounds in acute inflammation. Ann NY Acad Sci 1964;116:847.
- Zipf K, Geise W. Uber die wirkung adenosinartiger stoffe und einiger organextrakte auf die kapillaren. Arch Pharmakol Exp Pathol 1933;171:111.
- 26. Werle E, Gotze W, Keppler A. Uber die wirking des kallikreins auf den isolierten darm und uber eine neue darmkontrahierende substanz. Biochem Z 1937; 289:217.
- 27. Werle E, Berek U. Zur kenntnis des kallikreins. Angew Chem 1948;60A:53.
- 28. Werle E, Berek U. Uber kallidin. Biochem Z 1950;320:136.
- Spector WG. The role of some higher peptides in inflammation. J Pathol Bact 1951; 63:93.
- MacKay ME, Miles AA, Schachter M, Wilhelm DL. Susceptibility of the guinea pig to pharmacological factors from its own serum. Nature 1953:172:714.
- Miles AA, Wilhelm DL. Enzyme-like globulins from serum producing the vascular phenomena of inflammation. I. An active permeability factor and its inhibitor in guinea pig serum. Br J Exp Pathol 1955;36:71.
- 32. Spector WG. The mediation of altered capillary permeability in acute inflammation. J Pathol Bact 1956;72:367.
- Wilhelm DL, Miles AA, MacKay ME. Enzyme-like globulins from the serum reproducing the vascular phenomena of inflammation. II. Isolation and properties of the permeability factor and its inhibition. Br J Exp Pathol 1955;36:82.
- Moulton R, Spector WG, Willoughby DA. Histamine release and pain production by xanthosine and related compounds. Br J Pharmacol 1957;12:365.
- 35. Spector WG. Activation of a globulin system controlling capillary permeability in inflammation. J Pathol Bact 1957;74:67.
- Spector WG. Substances which affect capillary permeability. Pharmacol Rev 1958;10:475.

- 37. Wilhelm DL, Mill PJ, Miles AA. Enzyme-like globulins from serum reproducing the vascular phenomena of inflammation. III. Further observations on the permeability factor and its inhibitor in guinea pig serum. Br J Exp Pathol 1957;38:446.
- 38. Wilhelm DL, Mill PJ, Sparrow EM, MacKay ME, Miles AA. Enzyme-like globulins from serum reproducing the vascular phenomena of inflammation. IV. Activable permeability factor and its inhibitor in the serum of the rat and the rabbit. Br J Exp Pathol 1958;39:228.
- Hurley JV. An electron microscopic study of leukocytic emigration and vascular permeability in rat skin. Aust J Exp Biol Med Sci 1963;41:171.
- Hurley JV. Substances promoting leukocyte emigration. Ann NY Acad Sci 1964;116:918.
- 41. Florey HW. The transport of materials across the capillary wall. Q J Exp Physiol 1964;49:117.
- Florey JW, Grant LH. Leukocyte emigration from small blood vessels stimulated with ultraviolet light: An electron microscope study. J Pathol Bact 1961;82:13.
- Movat HZ, Fernando NVP. Acute inflammation. I. The earliest fine structural changes at the blood tissue barrier. Lab Invest 1963;12:895.
- 44. Arnold J. Uber das verhalten der wandungen der blutgefasse bei der emigration weisser blutkorper. Arch Pathol Anat 1875;62:487.
- 45. Arnold J. Uber die Kittsubstanz der endothelien. Arch Pathol Anat 1876;66:77.
- Hurley JV, Xeros N. Electron microscope observations on the emigration of leukocytes. Aust J Exp Biol Med Sci 1961;36;609.
- Marchesi VT. The stie of leukocyte emigration in inflammation. Q J Exp Physiol 1961;46;115.
- Marchesi VT, Florey HW. Electron microscopic observations on the emigration of leukocytes. Q J Exp Physiol 1960;45:343.
- Spector WG, Willougby DA. Capillary-permeability factors, nucleosides and histamine release. J Pathol Bact 1957;73:133.
- Williamson JR, Grisham JW. Electron microscopy of leukocyte margination and emigration in acute inflammation in dog pancreas. Am J Pathol 1961;39:239.
- Zweifach BW. An analysis of the inflammatory reaction through the response of the terminal vascular bed to microtrauma. In: Jasmin G, Robert A, eds. The mechanism of inflammation. Montreal: Acta, 1953.
- Rebuck JW, LoGrippo GA. Characteristics and interrelationships of the various cells in the RE cell, macrophage, lymphocyte and plasma cell series in man. Lab Invest 1961;10:1068.
- 53. Sokol RJ, Durrant TE, Hudson G. Scanning electron microscopy of the skin window cells of normal subjects. J Anat 1978;126:157.
- 54. Rebuck JW, Petz AJ, Riddle JM, Priest RJ, Lo Grippo GA. Human leukocytic functions in the tissues. In: Wolstenholme GEW, O'Connor M, eds. Biological activity of the leukocyte. Ciba Foundation Study Group No. 10. Boston: Little Brown, 1961:3-31.
- 55. Priest RJ, Rebuck JW, Havey GT. A new qualitative defect of leukocyte function in ulcerative colitis. Gastroenterology 1960;38:715.
- Paz RA, Spector WG. The mononuclear cell response to injury. J Pathol Bact 1962;84;85.
- Volkman A, Gowans JH. The reproduction of macrophages in the rat. Br J Exp Pathol 1965;46:50.
- Cronkite EP, Fliedler TM. Granulocytopoiesis. N Engl J Med 1964;270:1347.

- 59. Ebert RH, Florey HW. The extravascular development of the monocyte observed in vivo. Br J Exp Pathol 1939;20:342.
- 60. Ryan GB. The origin and sequence of the cells found in the acute inflammatory response. Aust J Exp Biol Med Sci 1967;45:149.
- 61. Metchnikoff E (1893). Lectures on the comparative pathology of inflammation. Rpt. New York: Dover Publishing, 1968.
- 62. Kolouch F, Jr. The lymphocyte in acute inflammation. Am J Pathol 1939; 15:413.
- 63. Townsend WA, Campbell B. The effects of roentgen rays on the inflammatory cells of the mouse and rabbit. Blood 1949;4:1346.
- Good RA. Experimental allergic brain inflammation, a morphological study. J Neuropath Exp Neurol 1950;9:78.
- Berman L. Lymphocytes and macrophages in vitro. Their activities in relation to functions of small lymphocytes. Lab Invest 1966;15:1084.
- 66. Bjorklund B, Bjorklund V, Lundstrom R, Eklund G, Nilsson L, Gronneberg R. Cytokinetics of the destruction of HEp-2 in vitro by lymphoid cells from subjects immunized with HeLa antigen and demonstration of a mechanism for biologic amplification of certain lymphoid cells by phase contrast, time-lapse cinemicrography and scanning electron microscopy. J Reticuloendothel Soc 1972; 11:29.
- 67. Riis P. The cytology of inflammatory exudate. Thesis. Copenhagen: Munksgaard, 1959.
- 68. Braunsteiner H. Physiologie and physiopathologie der weissen blutzellen. Thieme. Stuttgart, Germany, 1959.
- Braunsteiner H, Partan J, Thumb N. Function of the lymphocytes. JAMA 1957;164:1604.
- Braunsteiner H, Partan J, Thumb N. Studies of lymphocytic functions. Blood 1958;8:417.
- 71. Eitzman DV, Smith RT. The nonspecific inflammatory response in the newborn period. J Dis Child 1957;94:484.
- 72. Eitzman DV, Smith RT. The nonspecific inflammatory cycle in the neonatal infant. J Dis Child 1959;97:326.
- Krivit W, Good RA. Aldrich's syndrome (thrombocytopenia, eczema and infection in infants). Studies of the defense mechanisms. J Dis Child 1959;97:137.
- Page AR, Good RA. Studies on cyclic neutropenia. J Dis Child 1957;94:623.
- Biggar WD, Holmes B, Page AR, Dienard AS, L'Esperance PL, Good RA. Metabolic and functional studies of monocytes in congenital neutropenia. Br J Haematol 1974;28:233.
- 76. Boxer LA, Hedley-Whyte ET, Stossel TP. Neutrophil actin dysfunction and abnormal behavior. N Engl J Med 1974;291:1093.
- Rebuck JW, Daher ME, Monaghan EA. Timing of glycogen transfer to and lysosomal and peroxisomal activity in inflammatory mononuclears of man. J Reticuloendothel Soc 1968;5:559.
- 78. Kontras SB, Bodenbender JG. Studies of the inflammatory cycle in juvenile diabetics. Am J Dis Child 1968;116:130.
- Perillie PE, Nolan JP, Finch SC. Studies of the resistance to infection in diabetes mellitus: Local exudative cellular response. J Lab Clin Med 1962;59:1008.
- Rebuck JW, Riddle JM, Barth CL. Abnormal leukocytic functions in test lesions in paroxysmal nocturnal hemoglobinuria. Fed Proc 1962;21:73.
- 81. Clark RA, Kimball HR. Defective granulocyte chemotaxis in the Chediak-Higashi syndrome. J Clin Invest 1971;50:2645.

- 82. Clark RA, Kimball HR, Padgett GA. Granulocyte chemotaxis in the Chediak-Higashi syndrome of mink. Blood 1972;39:644.
- Rebuck JW, Anderson JA, Weiss L, Sweet LC. Abnormality of the cellular response in an infant with hyperreactivity to cow's milk. Proceedings, 10th RES meeting, Williamsburg, Va, 1973:79.
- 84. Gewurz H, Page AR, Pickering RJ, Good RA. Complement activity and inflammatory neutrophil exudation in man. Studies in patients with glomerulonephritis, essential hypocomplementemia and agammaglobulinemia. Int Arch Allergy 1967;32:64.
- O'Connell EJ, Enriquez P, Linman JW, Gleich GJ, McDuffie FC. Absence of activity of first component of complement in man: Association with thymic alymphoplasia and defective inflammatory response. J Lab Clin Med 1967;70:715.
- Brayton RG, Stokes PE, Schwartz MS, Louia DB. Effect of alcohol and various diseases on leukocyte mobilization, phagocytosis, and intracellular bacterial killing. N Engl J Med 1970;282:123.
- McCabe WP, Rebuck JW, Kelly AP, Jr, Ditmars DM, Jr. Cellular immune response of humans to pigskin. Plast Reconst Surg 1973;51:181.
- McCabe WP, Rebuck JW, Kelley AP, Jr, Ditmars DM, Jr. Leukocytic response as a monitor of immuno-depression in burn patients. Arch Surg 1973;106:155.
- 89. Mlczoch F, Kohout J. Das "Gebebsbild" bei verschiedenen erkrankugen. Eine anwendung der deckglasmethode in der klinik. Klin Wochenschr 1962;40:99.
- 90. Ward PA, Johnson KJ, Kreutzer DL. Regulatory dysfunction in leukotaxis. Am J Pathol 1977;88:701.
- Cohn ZA, Hirsch JG. The isolation and properties of the specific cytoplasmic granules of rabbit polymorphonuclear leukocytes. J Exp Med 1960;112:983.
- 92. Cohn ZA, Hirsch JG. The influence of phagocytosis on the intracellular distribution of granule-associated components of polymorphonuclear leukocytes. J Exp Med 1960;112:1015.
- 93. Rebuck JW, Barth CL, Petz AJ. New leukocytic dysfunction at the inflammatory site in Hegglin's, Hurler's and Pelger-Heut anomalous states. Fed Proc 1963;22:247.
- Catovsky D. Fagocitosis in vivo con el metodo de la ventana cutanea. Observaciones en el ser humano normal y en hemopatias. Arch Func Roux-Ocefa 1967;1:113.
- 95. Hessburg PC, Rebuck JW. Skin window responses in uveitis. Am J Ophthalmol 1966;62:648.
- 96. Jessop JD, Vernon-Roberts V, Harris J. Effects of gold salts and prednisolone on inflammatory cells. I. Phagocytic activity of macrophages and polymorphs in inflammatory exudates studied by a "skin window" technique in rheumatoid and control patients. Ann Rheum Dis 1973;32:294.
- 97. Johnson AJ, Rebuck JW, Knoll BF. The effect of autogenous urine on leukocytic defenses in man. II. The responses to varied concentrations of saline, urea, and urine. Invest Urol 1970;8:224.
- 98. Knoll BF, Johnson AJ, Pearce CF, Rebuck JW. True effects of autogenous urine on leukocytic defenses in man. Surg Forum 1966;17:517.
- 99. Knoll BF, Johnson AJ, Pearce CW, Rebuck JW. The effect of autogenous urine on leukocytic defenses in man. Invest Urol 1969;6:406.
- Rebuck JW, Sigler JW, Barth CL. Study of gouty tophi in skin windows in man. Fed Proc 1965;24:240.
- Stetson CA, Jr. Similarities in the mechanisms determining the Arthus and Schwartzmann phenomenon. J Exp Med 1951;94:347.

Leukocytes in Inflammation

- Cochrane CG, Weigle WO, Dixon FJ. The role of polymorphonuclear leukocytes in the initiation and cessation of arthus vasculitis. J Exp Med 1965;110:481.
- 103. Humphrey JH. The mechanism of the arthus reaction. I. The role of polymorphonuclear leukocytes and other factors in reversed passive arthus reactions in rabbits. Br J Exp Pathol 1955;36:268.
- Ward PA, Cochrane CG. Bound complement and immunologic injury of blood vessels. J Exp Med 1965;121:215.
- 105. Ward PA, Cochrane CG, Muller-Eberhard HJ. The role of serum complement in chemotaxis of leukocytes in vitro. J Exp Med 1965;122:327.
- Ward PA, Cochrane CG, Muller-Eberhard HJ. Further studies on the chemotactic factor of complement and its formation in vivo. Immunology 1966;11:141.
- 107. Jensen JA, Snyderman R, Mergenhagen SE. Chemotactic activity of guinea pig C'5-anaphylatoxin. In: Movat HZ, ed. Cellular and humoral mechanisms in anaphylaxis and allergy. New York: Karger, 1969.
- 108. Movat HZ, Fernando NVP, Uriuhara T, Weiser WJ. Allergic inflammation. III. The fine structure of collagen fibrils at sites of antigenantibody interaction in arthus type lesions. J Exp Med 1963:118:557.
- 109. Movat HZ, Macmorine DRL, Takeuchi Y, Burrowes CE. Chemical mediators released by PMN-leukocytes during phagocytosis of Ag-Ab complexes. In: Movat HZ, ed. Cellular and humoral mechanisms in anaphylaxis and allergy. New York: Karger, 1969.
- 110. Uriuhara T. Phagocytosis of antigen-antibody complexes. Fed Proc 1964;23:390 (abst).
- 111. Urihara T, Movat HZ. Allergic inflammation. IV. The vascular changes during the development and progression of the direct active and passive arthus reaction. Lab Invest 1964;13:1057.
- 112. Uriuhara T, Movat HZ. The role of PMN-leukocyte lysosomes in tissue injury, inflammation, hypersensitivity. I. The vascular changes and the role of PMN leukocytes in the reversed passive arthus reaction. Exp Mol Pathol 1966;5:539.
- Fowler JW, Lowell FC. The accumulation of eosinophils as an allergic response to allergen applied to the denuded skin surface. J Allergy 1966;37:19.
- 114. Clark RAF, Gallin JI, Kaplan AP. The selective eosinophil chemotactic activity of histamine. J Exp Med 1975;142:1462.
- 115. Kay AB, Austen KF. The IgE-mediated release of an eosinophil leukocyte chemotactic factor from human lung. J Immunol 1971;107:899.
- Kay AB, Stechschulte DJ, Austen KF. An eosinophil leukocyte chemotactic factor of anaphylaxis. J Exp Med 1970;133:602.

- 117. Gosh ML, Hudson G, Blackburn EK. Investigation of cutaneous inflammatory response in polycythemia vera. J Clin Pathol 1974;26:513 (abst).
- Waldmann R, et al. Fitzgerald factor: A hitherto unrecognized coagulation factor. Lancet 1975;1:949.
- Rebuck JW, Waldmann R, Abraham J, MacDonald R. Importance of contact activated coagulation factors in inflammation. J Reticuloendothel Soc 1976;20:58A.
- Bullock JD, Bodenbender JG. A simple laboratory aid in diagnosing food allergy. Ann Allergy 1970;28:127.
- 121. Priest RJ, Rebuck JW. The role of basophilic granulocytes in ulcerative colitis. Med Hygiene 1962;20:365.
- 122. Rebuck JW. New disease of basophilic leukocytes in man. Skin 1964;3:21.
- Rebuck JW, Hodson JM, Priest RJ, Barth CL. Basophilic granulocytes in inflammatory tissues of man. Ann NY Acad Sci 1963;103:409.
- Rebuck JW, Priest RJ, Kunz JL. Basophilic leukocytes in inflammatory exudates of man. Anat Rec 1960;136:263.
- Hu F, Fosnaugh RP, Bryan HG, Jacks D. Human skin window studies. II. Comparison of cellular response to staphylococcus in controls and in patients with cutaneous bacterial infections. J Invest Dermatol 1963;41:325.
- 126. Wolf-Jurgensen P. Dependence of the NLT-test upon quantitative and qualitative changes of the injected lymphocytes. Acta Pathol Microbiol Scand 1965;65:46.
- 127. Wolf-Jurgensen P. Basophilic leukocytes in delayed hypersensitivity. In: Experimental studies in man using the skin window technique. Copenhagen: Munksgaard, 1966.
- 128. Wolf-Jurgensen P, Schwartz M. Normal lymphocyte transfer in man. Basophil leukocytes in delayed skin reaction. Lancet 1964;2:388.
- 129. Scheie HG, Hambrick GW, Jr, Barness LA. A newly recognized forme fruste of Hurler's disease (gargoylism): The Sanford R. Gifford lecture. Am J Opthalmol 1962;53:573.
- Rebuck JW, Nixon RK, Manson G. Diagnostically abnormal mononuclear leukocytes in the spectrum of mucopolysaccharidoses. Fed Proc 1969;28:549.
- Rebuck JW, Schiller S. Histiocytic reactions to acid mucopolysaccharides in human skin windows. J Reticuloendothel Soc 1964;1:364.
- Kohout J. Macrophage reactivity in skin windows of sarcoidosis. Ann NY Acad Sci 1976;278:201.
- Marks JG, Thor DE, Loew RS. Darier's disease. An immunologic study. Arch Dermatol 1978;114:1336.